



## MasterTech ROUTINE H&E PROCEDURE

(Revised 02/20/18)

**PRINCIPLE:** This procedure stains cell nuclei and cytoplasm.

**SPECIMEN:** Any well fixed paraffin embedded tissue cut at 5 microns.

**QUALITY CONTROL:** American Master\*Tech Scientific Recommended Control Slide:  
Heart, CSH0725P; Skeletal Muscle, CSS0725P; Uterus, CSU0325P

**PROCEDURE NOTE:** If using a **Xylene Substitute**, deparaffinize for 10 minutes in each change with agitation; clear for 2 minutes in each change with agitation.

### **PROCEDURE:**

1. Deparaffinize slide in 2 changes of Xylene at 5 minutes each.
2. Place slide in 3 changes of Absolute Alcohol for 1 minute each with agitation.
3. Place slide in 95% Reagent Alcohol for 1 minute with agitation.
4. Rinse slide in Tap water for 1 minute.
5. Place slide in **MasterTech Harris Hematoxylin** for 5 minutes.
6. Rinse slide in Tap water for 1 minute.
7. Dip slide 2 to 3 times in **MasterTech Differentiating Solution**.
8. Rinse slide in Tap water for 1 minute.
9. Place slide in **MasterTech Bluing Solution** for 30 to 40 seconds.
10. Rinse slide in Tap water for 2 to 3 minutes.
11. Place slide in 70% Reagent Alcohol for 1 minute with agitation.
12. Place slide in **MasterTech Eosin Y Stain or Aqueous Eosin Y Stain** for 30 to 60 seconds with agitation.
13. Dehydrate slide through 4 changes of Absolute Alcohol for 1 minute each with agitation.
14. Clear slide through 3 changes of Xylene or Xylene Substitute for 1 minute each with agitation.
15. Coverslip using **ClearMount™**, **CoverSafe™**, or **MicroMount™**.

### **RESULTS:**

Nuclei: **BLUE**  
Cytoplasm: **PINK**

**REFERENCE:** Sheehan DC Hrapchak BB; Theory and Practice of Histotechnology; 1980, 143 - 144.

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