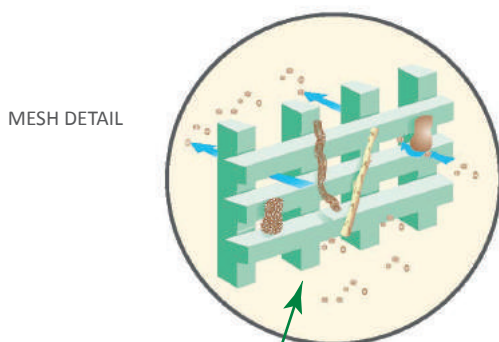
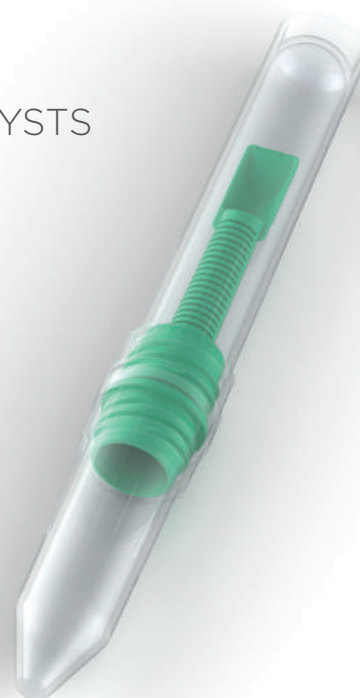


# FOR FAECAL CONCENTRATION OF

HELMINTH OVA AND LARVAE / PROTOZOA CYSTS AND OOCYSTS

APACOR

Mini Parasep®  
FAECAL PARASITE CONCENTRATOR



MESH DETAIL

SELF STANDING  
SAMPLE CHAMBER

MIXING CHAMBER

INTEGRAL  
SPOON

## Filter

A two stage filtration matrix. Large particles are rejected without occluding the 425µm pores. Recovery rate with Parasep® is comparable to traditional sieve method, ie: Ridley-Allen. The vertical filter enclosed design is patented.

## Debris Trap

Rejected particles are trapped to prevent extrusion into the Sedimentation Cone during centrifugation.

## Air/Liquid Seal and Safety Lock

The 'seal' prevents the release of biohazardous material. The 'lock' ensures the Mixing Chamber and Filter are removed together for safe disposal.

## Sedimentation Cone

Sediment forms in the base of the cone allowing examination for the presence of helminth eggs or larvae and protozoa cysts or oocysts.

## Health and Safety Benefits

- Totally enclosed/sealed process
- Reduced reagent volumes
- No cleaning required
- Single use, no sample contamination
- Ready to use systems available

## Performance Benefits

- Optimum sample recovery
- Enhanced sample clarity
- Rapid four step process
- Human resources optimised
- Easy patient identification
- Fits all 15ml centrifuge buckets



PARASITOLOGY

SINGLE USE IN VITRO DIAGNOSTIC DEVICE



FAECAL PARASITE CONCENTRATOR

Mini Parasep®

## Procedure

### STEP 1 - SAMPLE PREPARATION

For empty Parasep®, unscrew lid and add 2.4ml of 10% Formalin and one drop of surfactant (eg: Triton X-100) to the mixing chamber.

Alternatively use the reagent ready Mini Parasep®.

Introduce a pea sized faecal sample using the spoon on the end of the Mini Parasep® filter. Add 0.9ml of Ethyl Acetate to the mixing chamber.

Mix in thoroughly with the Mini Parasep® spoon. If the sample is hard, break it up with the end of the spoon.



### STEP 2 - EMULSIFICATION

Seal the Mini Parasep® by screwing in the filter/ sedimentation cone unit.

Vortex or shake to emulsify with the sedimentation cone pointing upwards.



### STEP 3 - CENTRIFUGATION

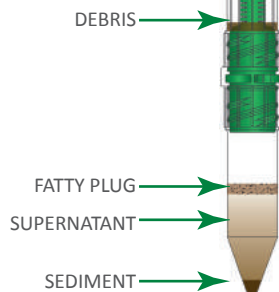
Invert Mini Parasep® and centrifuge at 1200g for three minutes.

Mini Parasep® fits all 15ml centrifuge buckets.

NOTE: TO CALCULATE THE REQUIRED RPM FOR ANY CENTRIFUGE.

$$RPM = \sqrt{\frac{g}{1.12r}} \times 1000$$

RPM - rotor speed in revs/min.  
g - centrifugal force (max. 1000g)  
r - radius, horizontal distance between sedimentation cone tip and spindle centre measured in mm.

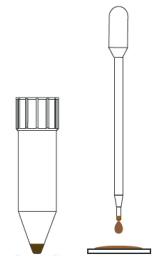


### STEP 4 - EXAMINATION

#### Direct Method

Unscrew and discard the filter and mixing tube. Pour off all the liquid above the sediment.

Pipette one drop of Saline or Iodine solution onto a slide, add one drop of deposit to the Saline or Iodine, mix sample and cover with cover-slip.

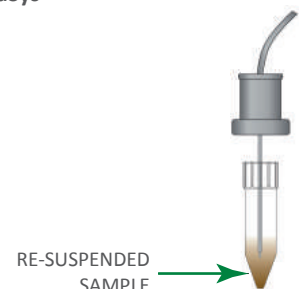


OR

#### Semi-automated System - ParaSys™

Unscrew and discard the filter and mixing tube. Pour off all the liquid above the sediment.

Press 'Dilute' to add Saline to the sediment. Shake or vortex to re-suspend sample. Insert Aspirator into suspension and press SAMPLE to draw 100µl into the ParaSlide™. (Refer to ParaSys™ instruction manual).



See label for storage conditions and expiry date. Please adhere to the following guidelines when handling Mini Parasep®. To avoid cross contamination the Mini Parasep® device should remain closed at all times except when introducing the sample or when retrieving the final concentrated sample for examination.

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