



## CELESTINE BLUE STAIN KIT PROCEDURE

Item# **CUKTCBL**

(Revised 03/7/18)

### KIT COMPONENTS INCLUDED:

100ml ZENKER'S FLUID W/MERCURIC CHLORIDE	100ml UNIVERSAL IODINE SOLUTION
100ml 5% SODIUM THIOSULFATE	100ml CELESTINE BLUE SOLUTION
100ml MODIFIED MAYER'S HEMATOXYLIN	200ml ORANGE G – PICRIC ACID
100ml 1% ACID FUCHSIN SOLUTION	100ml MACFARLANE'S STOCK SOLUTION
100ml 2% LIGHT GREEN SOLUTION	

**PRINCIPLE:** This kit demonstrates fibrin, keratin and cytoplasmic granules.

**SPECIMEN:** 10% Formalin fixed paraffin embedded tissue cut at 4 to 5 microns.

**QUALITY CONTROL:** American MasterTech Scientific Recommended Control Slide: Alzheimer's, CSA0225P

### MACFARLANE'S WORKING SOLUTION:

1. MACFARLANE'S STOCK SOLUTION ..... 40 ml
2. 95% Reagent Alcohol ..... 40 ml
3. Distilled water. .... 20 ml

### DIFFERENTIATING SOLUTION:

1. ORANGE G – PICRIC ACID SOLUTION ..... 30 ml
2. 80% Reagent Alcohol ..... 70 ml

### PROCEDURE:

1. Deparaffinize slide using Xylene or a Xylene Substitute and hydrate through graded alcohols as usual.
2. Rinse slide in Distilled water.
3. Place slide in **ZENKER'S FLUID WITH MERCURIC CHLORIDE** overnight.
4. Rinse slide thoroughly in running Tap water.
5. Place slide in **UNIVERSAL IODINE SOLUTION** for 5 minutes.
6. Rinse slide in running Tap water.
7. Place slide in **5% SODIUM THIOSULFATE** for 2 minutes.
8. Rinse slide in running Tap water for 15 minutes.
9. Place slide in **CELESTINE BLUE SOLUTION** for 5 minutes.
10. Rinse slide in running Tap water for 4 minutes.
11. Place slide in **MODIFIED MAYER'S HEMATOXYLIN** for 5 minutes.
12. Rinse slide in running Tap water for 5 minutes.
13. Place slide in **ORANGE G – PICRIC ACID SOLUTION** for 5 minutes.
14. Rinse slide in running Tap water for 1 minute.
15. Place slide in **1% ACID FUCHSIN SOLUTION** for 5 minutes.
16. Rinse slide in running Tap water for 1 minute.
17. Place slide in Differentiating Solution for 10 to 15 seconds.
18. Rinse slide in running Tap water for 1 minute.
19. Place slide in MacFarlane's Working Solution for 5 minutes.
20. Rinse slide in running Tap water for 1 minute.
21. Place slide in **2% LIGHT GREEN SOLUTION** for 1 minute.
22. Rinse slide in running Tap water for 15 seconds.
23. Dehydrate slide through 3 changes of 95% Reagent Alcohol.
24. Dehydrate slide through 3 changes of fresh Absolute Alcohol.
25. Clear slide through 3 changes of fresh Xylene or Xylene Substitute.
26. Coverslip using a permanent mounting media.

### RESULTS:

Fibrin and keratin: **RED**

Cytoplasmic granules: **RED**

Collagen: **GREEN**

Erythrocytes: **ORANGE**

**REFERENCES:** Lendrum AC, Fraser DS, Slidders W, Henderson R: J. Clin. Path. 15: 401–413, 1962; Luna LG: Histopathologic Methods and Color Atlas of Special Stain and Tissue Artifacts; American Histolabs, Inc.; 410 – 411, 1992

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