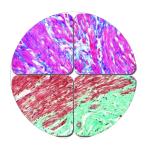
# **MasterTech** S<sup>2</sup>Stain Kit Instructions for Use



# **Trichrome One-Step (Green & Red)**

# Kit Item # QS2-TRIGR

Trichrome stains are used to identify increases in collagenous tissue to assist in the diagnosis of diseases such as liver cirrhosis. The One-Step Trichrome makes staining collagen or muscle fast and easy. One-step trichrome serves as the primary stain, producing pink to red cytoplasm, intercellular fiber, keratin, and muscle, and blue collagen and mucus.

INCLUDES COMPONENTS	Item#	Vials Included	
Dewax Solution	S001-15	1 vial (15mL)	
Bouin's Fluid	FXBOU-15	2 vials (15mL)	
Weigert's Hematoxylin A	HXWHEA-7	1 vial (7mL)	
Weigert's Hematoxylin B	HXWHEB-7	1 vial (7mL)	
One-Step Trichrome (Green & Red)	STOSTG-SF-15	1 vial (15mL)	
Reagent Alcohol, 100%	6900-15	1 vial (15mL)	

#### STORAGE AND STABILITY

Store components at room temperature. When properly stored, the reagents are stable to the date indicated on the label.

#### **RESULTS**

Cytoplasm: Pink to red Keratin: Pink to red Muscle: Pink to red

Intercellular fibers: Pink to red

Collagen: Green Mucus: Green Nucleus: Dark Blue

Each kit will stain approximately 50 slides. Intended for *in-vitro* use by laboratory professionals.

#### **SPECIMEN PREPARATION**

Appropriately fixed, paraffin-embedded, 3-5µm tissue section.

#### **CONTROL TISSUE**

Liver, heart, skeletal muscle, uterus, colon

### **DILUTION AND MIXING**

Most solutions in the kit are ready-to-use. The on-slide mixing feature on the Quantum S2 Slide Stainer will mix the working Weigert's Hematoxylin solution (Weigert's Hematoxylin (A) and Weigert's Hematoxylin (B).

#### LIMITATIONS AND PRECAUTIONS

See SDS for complete warnings, precautions, hazard and precautionary statements, and disposal information. Do not use if reagents become cloudy. Do not use past expiration date.

#### **INSTRUCTIONS FOR USE**

- 1 Press Prepare Labels to prepare slide labels, and affix labels to slides
- 2 Place and secure blue staining chambers in respective module lids.
- 3 Insert labeled slides on the modules and press Scan Slides.
- 4 Press **Scan Reagents** to display the required reagent names and volumes (number of tests).
- **5** Place Trichrome One-Step (Green & Red) Stain Kit vials onto the Reagent Rack and remove caps from vials.
- **6** Press **Scan Reagents** to start the staining process.

Use stains and reagents when they are at room temperature. Tissue section should be placed in proper area of microscope slide for best results. Refer to the tissue placement guide. Check the level of bulk deionized water before stain run to ensure proper volumes are used for optimal staining results. Replace caps on the vials when not using to minimize solution evaporation or other variables. The blue chambers must be cleaned after each use with Quantum Chamber Cleaning Solution for 20-30 minutes followed by a thorough deionized water rinse. Allow to air dry before each use.

#### **MATERIALS REQUIRED BUT NOT SUPPLIED**

- 1 Control tissue (CST0125P)
- 2 Blue Staining Chambers (QHD-CH200-10)
- **3** Quantum Chamber Cleaning Solution (QHD-QHS-1)
- 4 QS2 Cleaning Kit, Standard Special Stains (Alcohol) (QS2-CLN)

# **NOTES**

For possible customizations, staining protocol information, or troubleshooting, please contact the Technical Support Department at StatLab by emailing <a href="mailto:tech@StatLab.com">tech@StatLab.com</a> or calling 1-800-442-3573 ext. 106.



#### **STATLAB QUANTUM S2 STAINER**

Run more stains with the StatLab Quantum S2 Slide Stainer, a fully-automated slide staining system with the broadest stain portfolio available. This universal system is designed to automate the manual staining methods routinely used in special stains and related applications. Its user-friendly programming and flexible platform allow for easy user interface. The StatLab MasterTech S2 Stain Kits are to be used exclusively on the Quantum S2 Slide Stainer, and no other reagents should be used other than those provided in the kits or specified as they may damage the platform

# **REFERENCES**

- 1. Sheehan DC Hrapchak BB: Theory and Practice of Histotechnology; 1980, 190.
- 2. A.F.I.P. Laboratory Methods in Histotechnology: 1992, 132 133.
- 3. With modifications by AMTS R&D Department, 1979-2018.