MasterTech S² Stain Kit Instructions for Use



Picro-Sirius Red

Kit Item # QS2-PRS

The Picro-Sirius Red stain kit is used to identify collagen and muscle. This kit can be used in lieu of and comparable to Van Gieson's staining procedures. Picro-Sirius Red serves as the primary stain producing red collagen and yellow muscle. This stain can be viewed under a polarized light to demonstrate birefringence of both thick and thin fibers. Even without a polarized light, Picro-Sirius red stain demonstrates reticular fibers and basal lamina of cerebral capillaries, which are often not observed in a Van Gieson's stain and can be masked by other stained structures in trichrome procedures.

| INCLUDES COMPONENTS | ltem# | Vials Included | |
|-------------------------|-------------|----------------|--|
| Dewax Solution | S001-15 | 1 vial (15ml) | |
| Weigert's Hematoxylin A | HXWHEA-7 | 1 vial (7mL) | |
| Weigert's Hematoxylin B | HXWHEB-7 | 1 vial (7mL) | |
| Picro-Sirius Red | STPSR-SF-15 | 1 vial (15mL) | |
| Acetic Acid, 0.5% | KC671-15 | 1 vial (15mL) | |
| Reagent Alcohol, 100% | 6900-15 | 1 vial (15mL) | |
| | | | |

STORAGE AND STABILITY

Store components at room temperature. When properly stored, the reagents are stable to the date indicated on the label.

RESULTS

Collagen: Red Muscle: Yellow

Thick fibers: Yellow to orange birefringence when viewed using a polarized light

Thin fibers: Green birefringence when viewed using a polarized

light

Nuclei: Black

Intended for *in-vitro* use by laboratory professionals.

Each kit will stain approximately 50 slides.

SPECIMEN PREPARATION

Appropriately fixed, paraffin-embedded, 3-5µm tissue section.

CONTROL TISSUE

Uterus

DILUTION AND MIXING

All solutions in the kit are ready-to-use. No further dilution is required.

LIMITATIONS AND PRECAUTIONS

See SDS for complete warnings, precautions, hazard and precautionary statements, and disposal information. Do not use if reagents become cloudy. Do not use past expiration date.

INSTRUCTIONS FOR USE

- 1 Press **Prepare Labels** to prepare slide labels, and affix labels to slides
- 2 Place and secure blue staining chambers in respective module lids.
- 3 Insert labeled slides on the modules and press **Scan Slides**.
- **4** Press **Scan Reagents** to display the required reagent names and volumes (number of tests).
- 5 Place Picro-Sirius Red Stain Kit vials onto the Reagent Rack and remove caps from vials.
- **6** Press **Scan Reagents** to start the staining process.

Use stains and reagents when they are at room temperature. Tissue sections should be placed in proper area of the microscope slide for best results. Check the level of bulk deionized water before stain run to ensure proper volumes are used for optimal staining results. Replace caps on the vials when not using to minimize solution evaporation or other variables. The blue chambers must be cleaned after each use with Quantum Chamber Cleaning Solution for 20-30 minutes followed by a thorough deionized water rinse. Allow to air dry before each use.

MATERIALS REQUIRED BUT NOT SUPPLIED

- 1 Control tissue (CSU0325P)
- **2** Blue Staining Chambers (QHD-CH200-10)
- 3 QS2 Cleaning Kit, Standard Special Stains (Alcohol) (QS2-CLN)
- 4 Quantum Chamber Cleaning Solution (QHD-QCS-1)



NOTES

For possible customizations, staining protocol information, or troubleshooting, please contact the Technical Support Department at StatLab by emailing tech@StatLab.com or calling 1-800-442-3573 ext 106

STATLAB QUANTUM S2 STAINER

Run more stains with the StatLab Quantum S2 Slide Stainer, a fully-automated slide staining system with the broadest stain portfolio available. This universal system is designed to automate the manual staining methods routinely used in special stains and related applications. Its user-friendly programming and flexible platform allow for easy user interface. The StatLab MasterTech S2 Stain Kits are to be used exclusively on the Quantum S2 Slide Stainer, and no other reagents should be used other than those provided in the kits or specified as they may damage the platform.

REFERENCES

- 1. Sheehan DC Hrapchak BB: Theory and Practice of Histotechnology; 1980, 190.
- 2. A.F.I.P. Laboratory Methods in Histotechnology: 1992, 132 133.
- 3. With modifications by AMTS R&D Department, 1979-2018.