Giemsa Stain Kit (May-Grunwald) (For Bone Marrow)

Description: The Giemsa Stain Kit (May-Grunwald) is intended for use in the visualization of cells present in hematopoietic tissues and certain microorganisms. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Uses/Limitations: Not to be taken internally. For In-Vitro Diagnostic use only. Histological applications. Do not use if reagents become cloudy. Do not use past expiration date. Use caution when handling reagents. Non-Sterile.

Results:

<table>
<thead>
<tr>
<th>Item</th>
<th>Kit Contents</th>
<th>Volume</th>
<th>Storage</th>
</tr>
</thead>
<tbody>
<tr>
<td>SSC-MAY500</td>
<td>May-Grunwald Stock Solution</td>
<td>500 ml</td>
<td>18-25°C</td>
</tr>
<tr>
<td>SSC-GGS500</td>
<td>Giemsa Stock Solution</td>
<td>500 ml</td>
<td>18-25°C</td>
</tr>
<tr>
<td>SSC-PBM500</td>
<td>Phosphate Buffer Solution, pH 6.8</td>
<td>500 ml</td>
<td>18-25°C</td>
</tr>
</tbody>
</table>

Precautions: Keep away from open flame. Avoid contact with skin and eyes. Harmful if swallowed. Follow all Federal, State, and local regulations regarding disposal. Use in chemical fume hood whenever possible.

Note: Individual components are designed to be interchangeable with StatLab kits when both are produced by StatLab and have identical catalog numbers (e.g. SSC-Mayxxx may be ordered as an individual component to replace May Grunwald Stock Solution that is supplied with kit.)

Preparation of Reagents Prior to Beginning:


2. Prepare **Working Giemsa Solution** by mixing 2.5ml of Giemsa Stock Solution (GGS500) with 50ml of Phosphate Buffer Solution, pH 6.8.

Procedure (Standard):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide in staining tray and flood with Working May-Grunwald Solution for 5-7 minutes.  
   Note: Agitate slide occasionally to insure proper staining.
3. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
   Note: Agitate slide occasionally to insure proper staining.
5. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
6. Allow slide to remain in Phosphate Buffer Solution, pH 6.8 for an additional 3 minutes.
7. Dip slide quickly in distilled water to remove buffer and air dry at room temperature.
8. Dip slide twice in Xylene or Xylene Substitute.

Procedure (Mast Cells):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide in staining tray and flood with Working May-Grunwald Solution for 5-7 minutes.  
   Note: Agitate slide occasionally to insure proper staining.
3. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
   Note: Agitate slide occasionally to insure proper staining.
5. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
6. Differentiate by dipping slide in Acetic Acid Solution (0.25%) until background is desired intensity.
7. Dip slide for 10 seconds in Phosphate Buffer Solution, pH 6.8 while agitating gently.
8. Dip slide quickly in distilled water to remove buffer and air dry at room temperature.
9. Dip slide twice in Xylene or Xylene Substitute.
10. Mount in synthetic resin.
References: